

Unfortunately I cannot comment on MetaprobeTM biomarkers since I have not managed to find any information on their use. However, the following comments may be relevant:

Liver function

Cytochrome P4501A2, the principal liver enzyme involved in aromatic amine N-oxidation, exhibits a wide range of interindividual variation for which various phenotyping methods using the probe drug caffeine exist [1]. These methods include the determination of various caffeine metabolite ratios in saliva [2,3] and urine [3-5], and caffeine-derived labeled CO in exhaled breath [6]. Dietary caffeine intake [2], smoking [2,4,5,7,8], gender [8], use of oral contraceptives [2] and chronic alcohol consumption [7] are associated with increased caffeine clearance (i.e., CYP1A2 induction). Furthermore, a C→A point mutation polymorphism in intron 1 of the CYP1A2 gene appears to confer high inducibility [3]. Thus, a gene-environment interaction may be a significant cause of the reported induction of CYP1A2 activity.

Contrary to the suggestion by Parexel, this test should not be used to identify 'closet smokers' since it lacks both sensitivity and specificity, and the study already includes determination of nicotine metabolites which will identify misreporting of smoking status.

Immune capacity

I could not find any information on the Tylenol challenge test or on other breath tests for measuring oxidative stress.

Mitochondrial capacity

The suggested assessment of mitochondrial capacity [9] uses administration of labeled ketoisocaproate and 2 hours post-administration determination of the decarboxylated analogue in exhaled breath. Studies of this kind normally use sequestered subjects and are therefore not ideally suited for the TE study. Since the effect of smoking has not previously been investigated it might be premature to use this type of breath test in the TE study.

Vascular epithelial health

Nitric oxide production, a non-specific marker of inflammatory response, can be measured in exhaled breath [10]. Although exhaled NO is easy to determine using a nitric oxide monitor, standardization of exhaled breath collection is difficult. Alternative methods for indirect determination of NO have focused mainly on measurement of NO metabolites (nitrate and nitrite) in induced sputum supernatant [11,14] and blood plasma [11]; however, these methods are not specific. Other methods, which are also not specific, include the determination of NO-related amino acids L-arginine and L-citrulline, and the intermediate N^ω-hydroxy-L-arginine in plasma [13].

Other metaprobes

I think these are outside our field of interest.

Technical suggestions:

3-Ethenyl-pyridines

3-Ethenylpyridine is a useful marker for indoor air ETS gas phase monitoring. It cannot be used as a biomarker of smoke exposure since its metabolism in man (and experimental animals) has not been established. By analogy to other compounds of similar structure it would be predicted to undergo epoxidation in the side chain and hydrolysis of the epoxide to the diol. However, a number of compounds in tobacco smoke with a pyridine moiety and an alkyl group at the 3 position could undergo biotransformation to give similar products. Therefore, 3-ethenylpyridine cannot be recommended as a potential biomarker of tobacco smoke exposure.

Hb adducts of 3- and 4-aminobiphenyl

The hemoglobin adducts of 3-aminobiphenyl and 4-aminobiphenyl have been extensively reported as suitable biomarkers of exposure in both smokers and nonsmokers [14-17]. 4-Aminobiphenyl hemoglobin adduct levels fall on smoking cessation with an estimated half-life of 7-9 weeks, consistent with the lifetime of hemoglobin [17]. However, two technical limitations have to be taken into consideration: (1) 4-aminobiphenyl hemoglobin adduct levels differ significantly between slow and rapid acetylators [14,16], and (2) 4-aminobiphenyl DNA adduct levels in human peripheral lung do not correlate with smoking status [18].

Nicotine and nicotine metabolites

Determination of nicotine, cotinine, 3'-hydroxycotinine and their respective glucuronide conjugates in 24-h urine samples will give a better estimation of nicotine exposure than determination of nicotine, cotinine and 3'-hydroxycotinine in spot blood samples. The presence of glucuronide conjugates of nicotine and its metabolites have not been reported in blood.

Thromboxane B2 and Fibrinogen

This is something where we do need some external advice on suitable biomarkers of effect.

Isoprostanes

Determination of isoprostanes as biomarkers of effect has on numerous occasions been recommended by Benowitz; however, I have yet to see any data from Benowitz on such studies. I agree with Parexel that a single isoprostane probably needs to be considered, and am currently in the process of reviewing the literature to identify the most suitable candidate biomarker(s). The F2-isoprostane family consists of a series of chemically stable prostaglandin F2 derivatives formed by peroxidation of arachidonic acid in phospholipids. Consequently, these compounds are suggested to represent a reliable index of lipid peroxidation (oxidative stress) in vivo [19]. However, analysis is both time consuming and complex. Recently a simplified method was reported using solid-phase extraction columns, high-performance liquid chromatography, and stable isotope dilution capillary gas chromatography/electron capture negative ionization mass spectrometry for 8-epi-PGF2 α , one of the most abundant F2-isoprostanes in biological samples [20]. I cannot make any more specific comments until I complete review of the literature on smoking and isoprostane levels.

Isopentane is not generally determined in exhaled air; however, pentane in exhalate has been determined as an index of lipid peroxidation [21,22]. Although

hydrocarbon-based breath tests for determination of oxidative stress are noninvasive, several minor technical difficulties (e.g., standardized washout period, air contamination) need to be resolved [23]. Without a complete review of the literature (over 100 publications on pentane in exhaled breath) I cannot make any further comments. However, my initial impression is that real-time monitoring of exhalate is probably not an area of biomonitoring that we want to get involved in.

Bronchial cells

I agree with Parexel that collecting nasal lavage may be technically less difficult than collecting induced sputum. However, the question still remains as to what should be investigated in the collected samples. Clara cell protein-16 (CC-16), a biomarker suggested by HHO, can still be measured in nasal lavages [5], but I am unsure as to what additional information this marker will contribute to the TE study.

Collecting induced sputum has the advantage that previous molecular epidemiology studies have investigated smoking-associated DNA adduct formation in bronchoalveolar macrophages [25,26]. However, it is unlikely that the required careful standardization of induced sputum collection using saline washing [27] was applied in these studies.

- 1 Fuhr U, and RL Rost (1994). Simple and reliable CYP1A2 phenotyping by the paraxanthine/caffeine ratio in plasma and in saliva. *Pharmacogenetics* 4 109-116.
- 2 Trantcheva-Poor I, M Zeigler, S Rietbrock, and U Fuhr (1999). Estimation of cytochrome P-450 CYP1A2 activity in 863 healthy Caucasians using a saliva-based caffeine test. *Pharmacogenetics* 9, 131-144.
- 3 Sachse C, J Brockmoller, S Bauer, and I Roots (1999). Functional significance of a C→A polymorphism in intron 1 of the cytochrome P450 CYP1A2 gene tested with caffeine. *Br. J. Clin. Pharmacol.*, 47, 445-449.
- 4 Nordmark A, S Lundgren, S Cnattinguis, and A Rane (1999). Dietary caffeine as a probe for assessment of cytochrome P4501A2 activity in random urine samples. *Br. J. Clin. Pharmacol.*, 47, 397-402.
- 5 Kalow W, and BK Tang (1991). Caffeine as a metabolic probe: exploration of the enzyme-inducing effect of cigarette smoking. *Clin. Pharmacol. Ther.*, 49, 44-48.
- 6 Parker AC, T Preston, D Heaf, NR Kitteringham, and I Choonara (1994). Inhibition of caffeine metabolism by ciprofloxacin in children with cystic fibrosis as measured by the caffeine breath test. *Br. J. Clin. Pharmacol.*, 38, 573-576.
- 7 Rizzo N, E Hispard, S Dolbeault, S Dally, R Leverage, and C Girre (1997). Impact of long-term ethanol consumption on CYP1A2 activity. *Clin. Pharmacol. Ther.*, 62, 505-509.
- 8 Schrenk D, D Brockmeier, K Mörike, KW Bock, and M Eichelbaum (1998). A distribution study of CYP1A2 phenotypes among smokers and non-smokers in a cohort of healthy Caucasian volunteers. *Eur. J. Clin. Pharmacol.*, 53, 361-367.
- 9 Lauterburg BH, I Grattagliano, R Gmur, M Stalder, and P Hildebrand (1995). Noninvasive assessment of the effect of xenobiotics on mitochondrial function in human beings: studies with acetylsalicylic acid and ethanol with the use of the carbon-13 labeled ketoisocaproate breath test. *J. Lab. Clin. Med.*, 125, 378-383.
- 10 Silkoff PE (2000). Noninvasive measurement of airway inflammation using exhaled nitric oxide and induced sputum. Current status and future use. *Clin. Chest Med.*, 21, 345-360.

- 11 Jang AS, and IS Choi (1999). Nitric oxide metabolites in patients with asthma; induced sputum versus blood. *Respir. Med.*, 93, 912-918.
- 12 Rutgers SR, TW van der Mark, W Coers, H Moshage, W Timens, HF Kauffman, GH Koeter, and DS Postma (1999). Markers of nitric oxide metabolism in sputum and exhaled air are not increased in chronic obstructive pulmonary disease. *Thorax* 54, 576-580.
- 13 Garlachs CD, J Beyer, H Zhang, A Schmeisser, K Plotze, A Mugge, S Schellong, and WG Daniel (2000). Decreased plasma concentrations of L-hydroxy-arginine as a marker of reduced NO formation in patients with combined cardiovascular risk factors. *J. Lab. Clin. Med.*, 135, 419-425.
- 14 Riffleman M, G Müller, W Schmieding, W Popp, and K Norpoth (1995). Biomonitoring of urinary aromatic amines and arylamine hemoglobin adducts in exposed workers and nonexposed control persons. *Int. Arch. Occup. Environ. Health*, 68, 36-43.
- 15 Bryant MS, PL Skipper, SR Tannenbaum, and M Maclure (1988). Hemoglobin adducts of 4-aminobiphenyl in smokers and nonsmokers. *Cancer Res.*, 47, 602-608.
- 16 Y MC, PL Skipper, K Taghizadeh, SR Tannenbaum, KK Chan, BE Henderson, and RK Ross (1994). Acetylator phenotype, aminodiphenyl-hemoglobin adduct levels, and bladder cancer risk in white, black, and asian men in Los Angeles, California. *J. Natl. Cancer Inst.*, 86, 712-716.
- 17 Mooney LA, RM Santella, L Covey, AM Jeffrey, W Bigbee, MC Randall, TB Cooper, R Ottman, WY Tsai, L Wazneh, AH Glassman, TL Young, and FP Perera (1995). Decline of DNA damage and other biomarkers in peripheral blood following smoking cessation. *Cancer Epidemiol. Biomarkers & Prev.*, 4, 627-634.
- 18 Culp SJ, DW Roberts, G Talaska, NP Lang, PP Fu, JO Lay, CH Teitel, JE Snawder, LS Von Tungeln, and FF Kadlubar (1997). Immunochemical, ³²P-postlabeling, and GC/MS detection of 4-aminobiphenyl-DNA adducts in human peripheral lung in relation to metabolic activation pathways involving pulmonary N-oxidation, conjugation, and peroxidation. *Mutation Res.*, 378, 97-112.
- 19 Lawson JA, J Rokach, and GA FitzGerald (1999). Isoprostanes: formation, analysis and use as indices of lipid peroxidation in vivo. *J. Biol. Chem.*, 274, 24441-24444.
- 20 Mori TA, KD Croft, IB Puddey, and LJ Beilin (1999). An improved method for measurement of urinary and plasma F2-isoprostanes using gas chromatography-mass spectrometry. *Anal. Biochem.*, 268, 117-125.
- 21 Mitsui T, K Naitoh, T Tsuda, T Hirabayashi, and T Kondo (2000). Is endogenous isoprene the only coeluting compound in the measurement of breath pentane? *Clin. Chim. Acta.*, 299, 193-198.
- 22 Mohler ER, P Reaven, JE Stegner, NS Fineberg, and DR Hathaway (1996). Gas chromatographic method using photoionization detection for the determination of breath pentane. *J. Chromatogr. B. Biomed. Appl.*, 685, 201-209.
- 23 Kneepkens CM, C Ferreira, G Lepage, and CC Roy (1992). The hydrocarbon breath test in the study of lipid peroxidation: principles and practice. *Clin. Invest. Med.*, 15, 163-186.
- 24 Lindahl M, J Svartz, and C Tagesson (1999). Demonstration of different forms of the anti-inflammatory proteins lipocortin-1 and Clara cell protein-16 in human nasal and bronchoalveolar lavage fluids. *Electrophoresis*, 20, 881-890.
- 25 Nia AB, LM Mass, SG Van Breda, DM Curfs, JC Kleinjans, EF Wouters, and FJ Van Schooten (2000). Application of induced sputum for molecular dosimetry of

- exposure to inhalatory carcinogens: ^{32}P -postlabeling of lipophilic DNA adducts in smokers and nonsmokers. *Cancer Epidemiol. Biomarkers & Prev.*, 9, 367-372.
- 26 Godschalk RW, LM Mass, N Van Zandwijk, LJ van't Veer, A Breedijk, PJ Borm, J Verhaert, JC Kleinjans, and FJ Schooten (1998). Differences in aromatic-DNA adduct levels between alveolar macrophages and subpopulations of white blood cells from smokers. *Carcinogenesis* 19, 819-825.
- 27 Gershman NH, H Liu, HH Wong, JT Liu, and JV Fahy (1999). Fractional analysis of sequential induced sputum samples during sputum induction: evidence that differential lung compartments are sampled at different time points. *J. Allergy Clin. Immunol.*, 104, 322-328.